

## Appendix D

### Supplementary Information for Chapter 5: Argentination: a silver bullet for cannabinoid analysis by DMS-MS<sup>2</sup>

This appendix contains the supporting information for Chapter 5, and is analogous to the supporting information to the following manuscript:

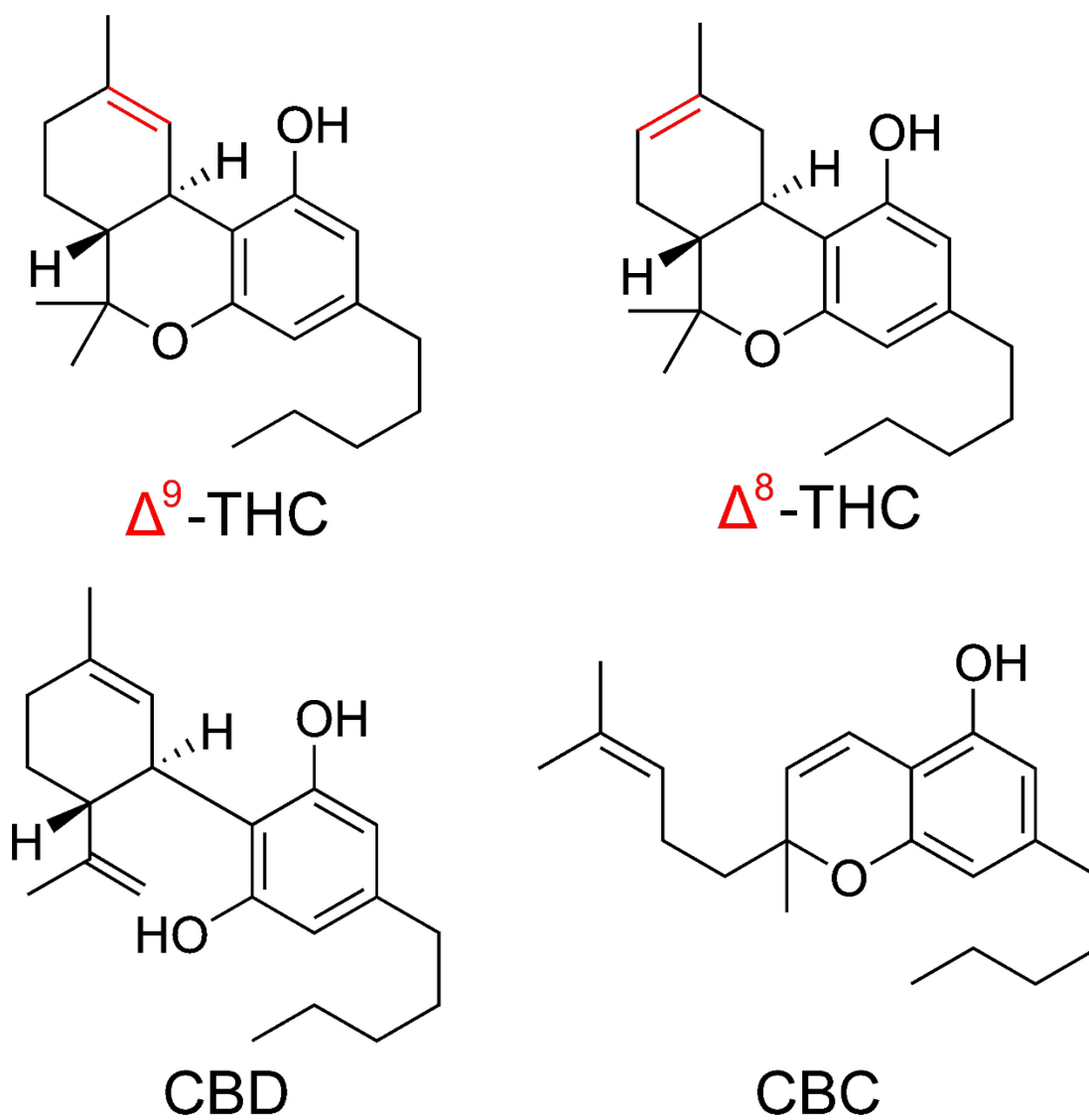
Ieritano, C., Thomas, P., Hopkins, W. S. Argentination: A Silver Bullet for Cannabinoid Analysis by Differential Mobility Spectrometry. *Anal. Chem.* **2023**. 95 (22), 8668 – 8678.

<https://pubs.acs.org/doi/abs/10.1021/acs.analchem.3c01241>

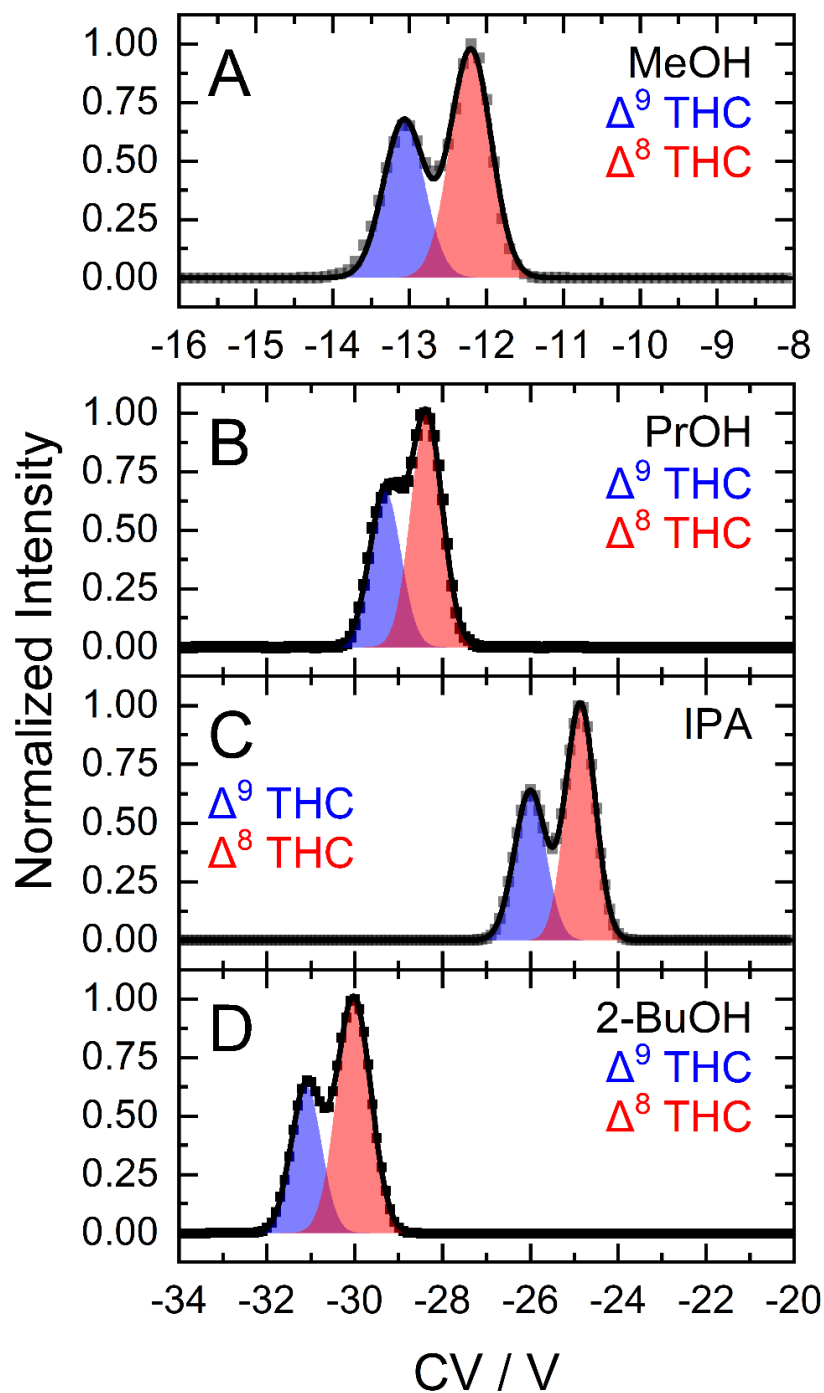
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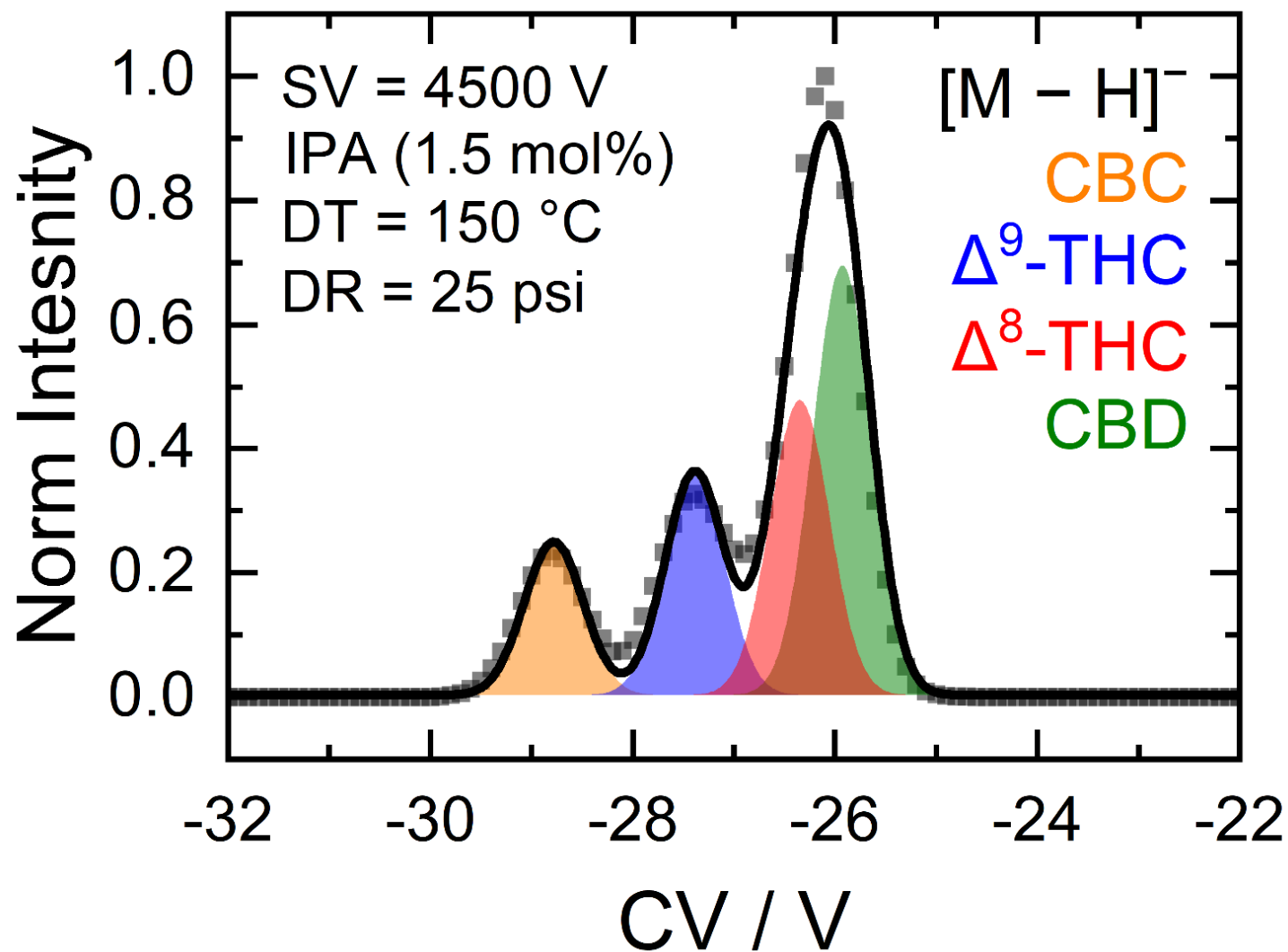
## Supporting Figures and Tables



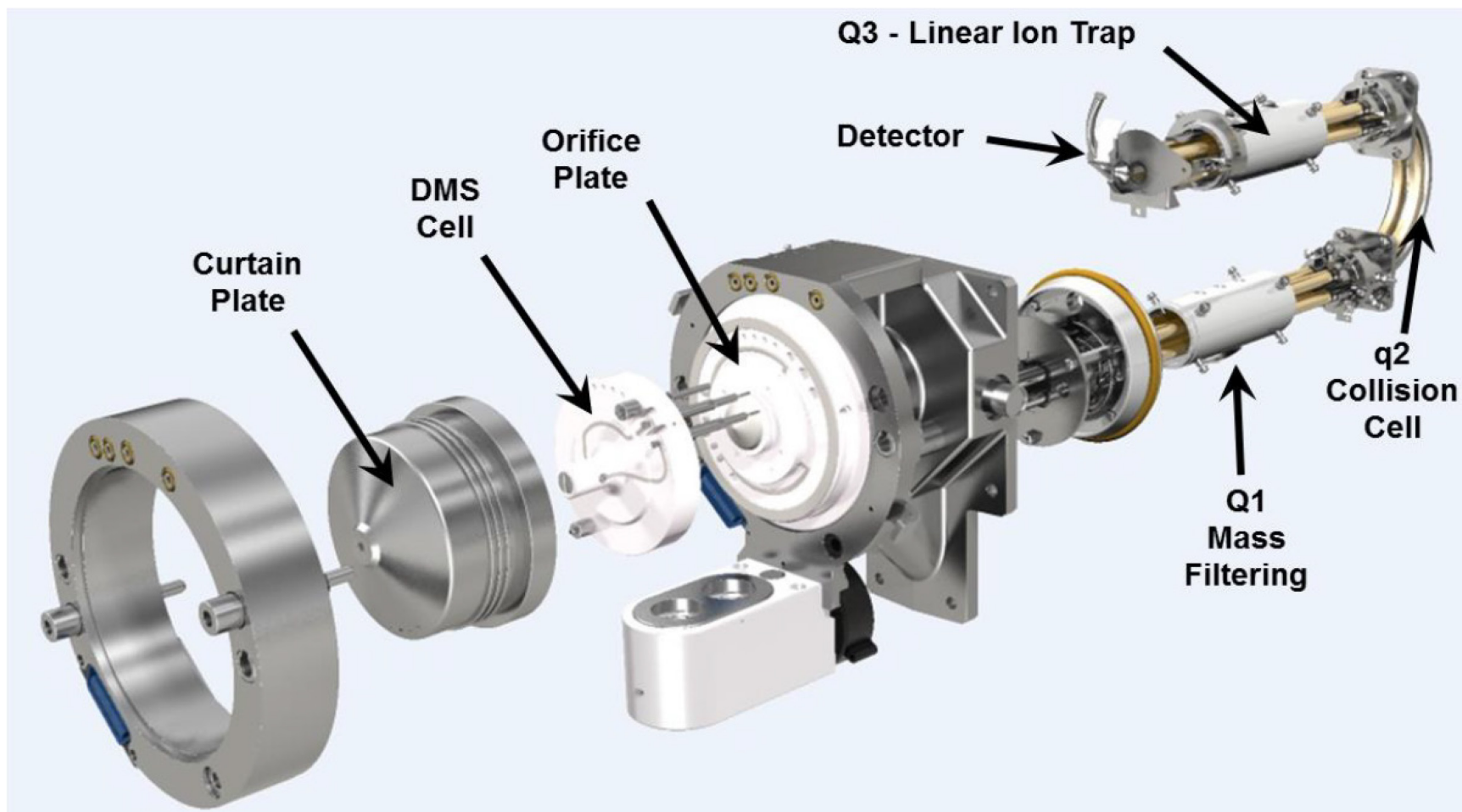
**Figure S5-1.** Structures of  $\Delta^9$ -THC,  $\Delta^8$ -THC, CBD, and CBC. Differences in double bond position between  $\Delta^8$ / $\Delta^9$ -THC are highlighted in red.



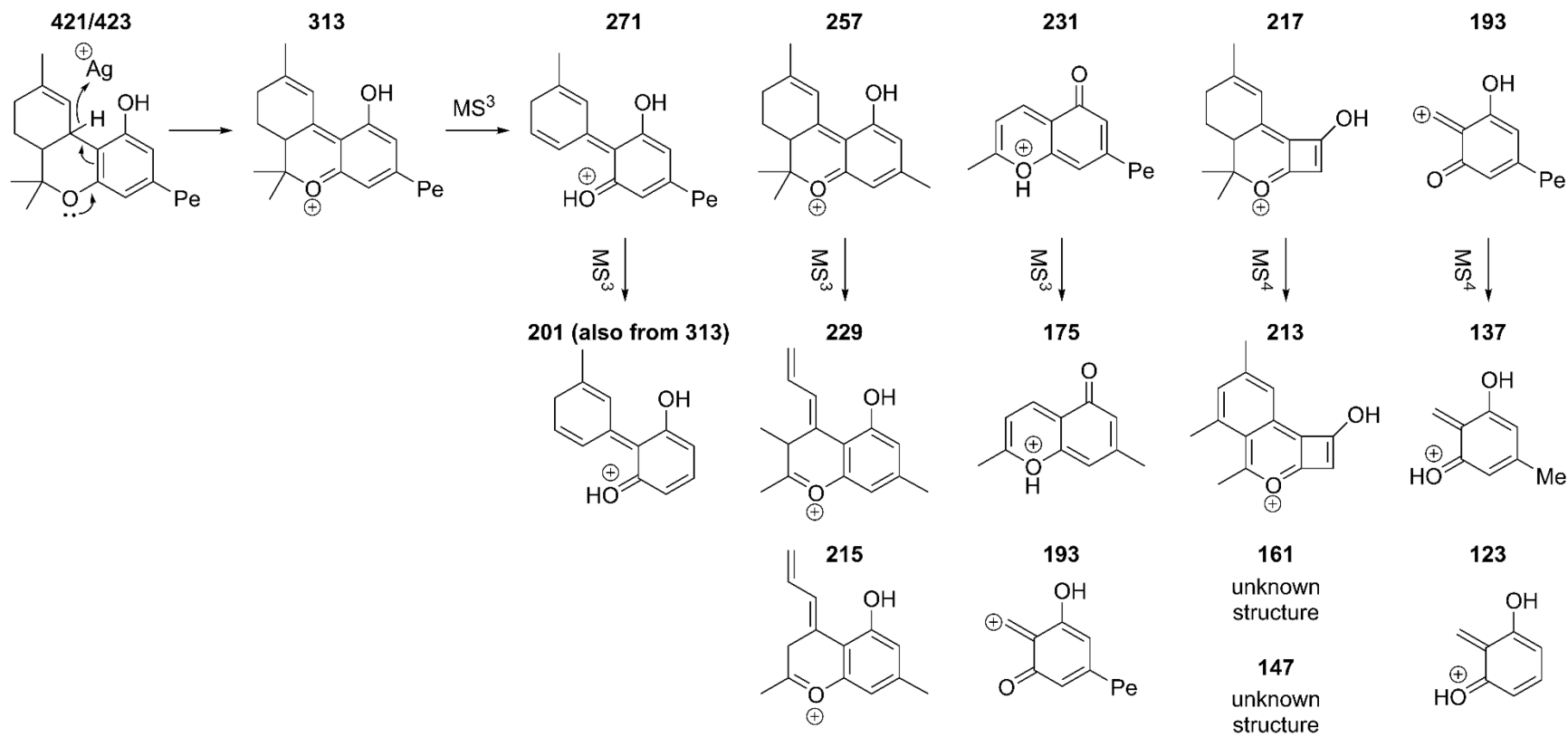
**Figure S5-2.** Ionograms of a 1:1 mixture of  $\Delta^9$ -THC and  $\Delta^8$ -THC. Analytes were monitored as deprotonated ions ( $[M - H]^-$ ;  $m/z$  313) in an  $N_2$  DMS environment seeded with 1.5 mol% of (A) MeOH, (B) 1-propanol, (C) IPA, and (D) 2-butanol at  $SV = 4500$  V and  $DT = 150$  °C. 25 psi of resolving gas (DR gas;  $N_2$ ) was introduced to improve resolving power.



**Figure S5-3.** Ionograms of a 1:1:1:1 mixture of CBC,  $\Delta^9$ -THC,  $\Delta^8$ -THC, and CBD. Analytes were monitored as deprotonated ions ( $[M - H]^-$ ;  $m/z$  313) in an  $N_2$  DMS environment seeded with 1.5 mol% of IPA at SV = 4500 V and DT = 150 °C. 25 psi of resolving gas (DR gas;  $N_2$ ) was introduced to improve resolving power.



**Figure S5-4.** Schematic of the SELEXION system coupled to the QTRAP 5500 (SCIEX) hybrid linear ion trap triple-quadrupole mass spectrometer.



**Figure S5-5.** Proposed fragmentation pathway for  $[\Delta^9\text{-THC} + \text{Ag}]^+$ .

Plausible mechanisms of formation:

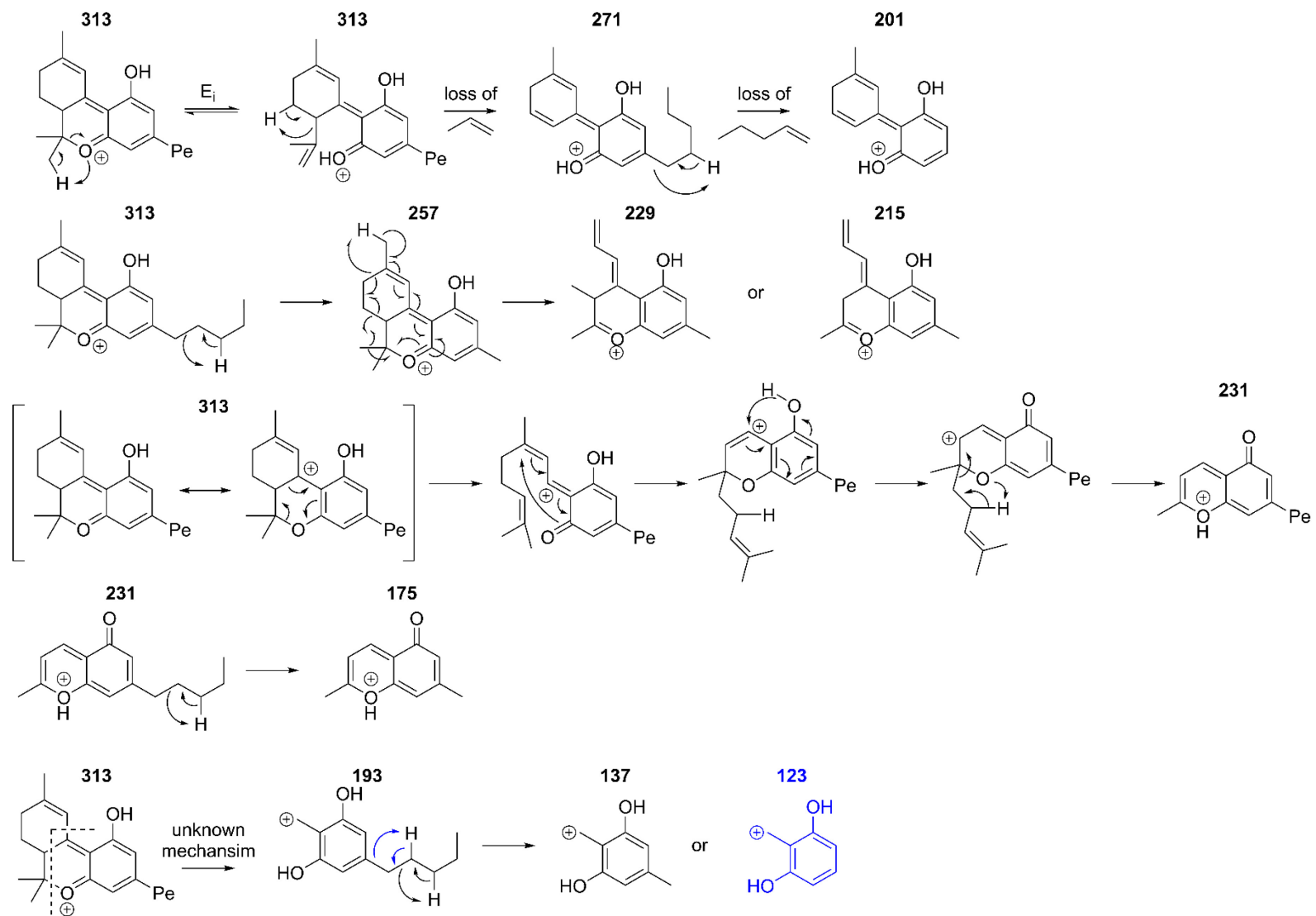
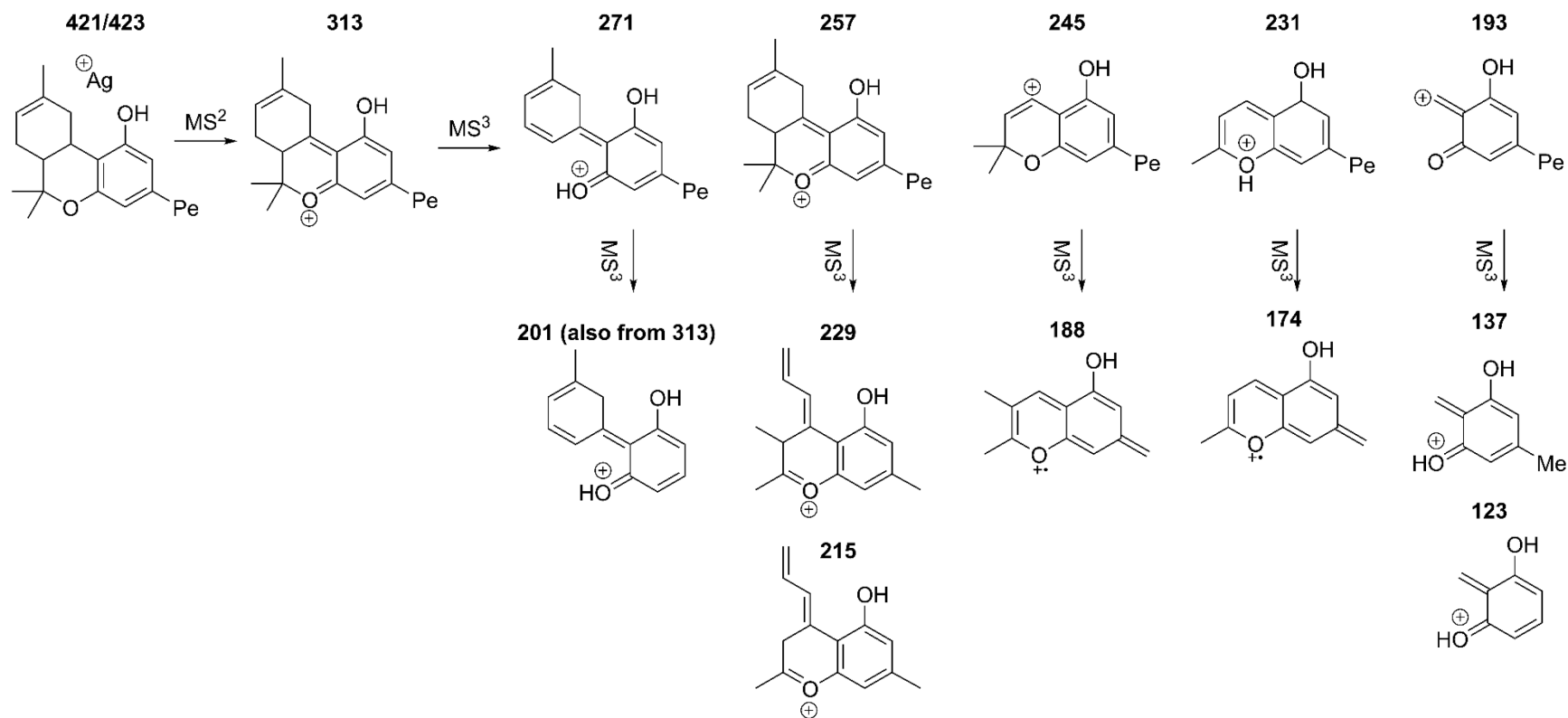


Figure S5-6. Proposed mechanisms of formation for fragments originating from  $[\Delta^9\text{-THC} + \text{Ag}]^+$ .





**Figure S5-7.** Proposed fragmentation pathway for  $[\Delta^8\text{-THC} + \text{Ag}]^+$ .

Plausible mechanisms of formation:

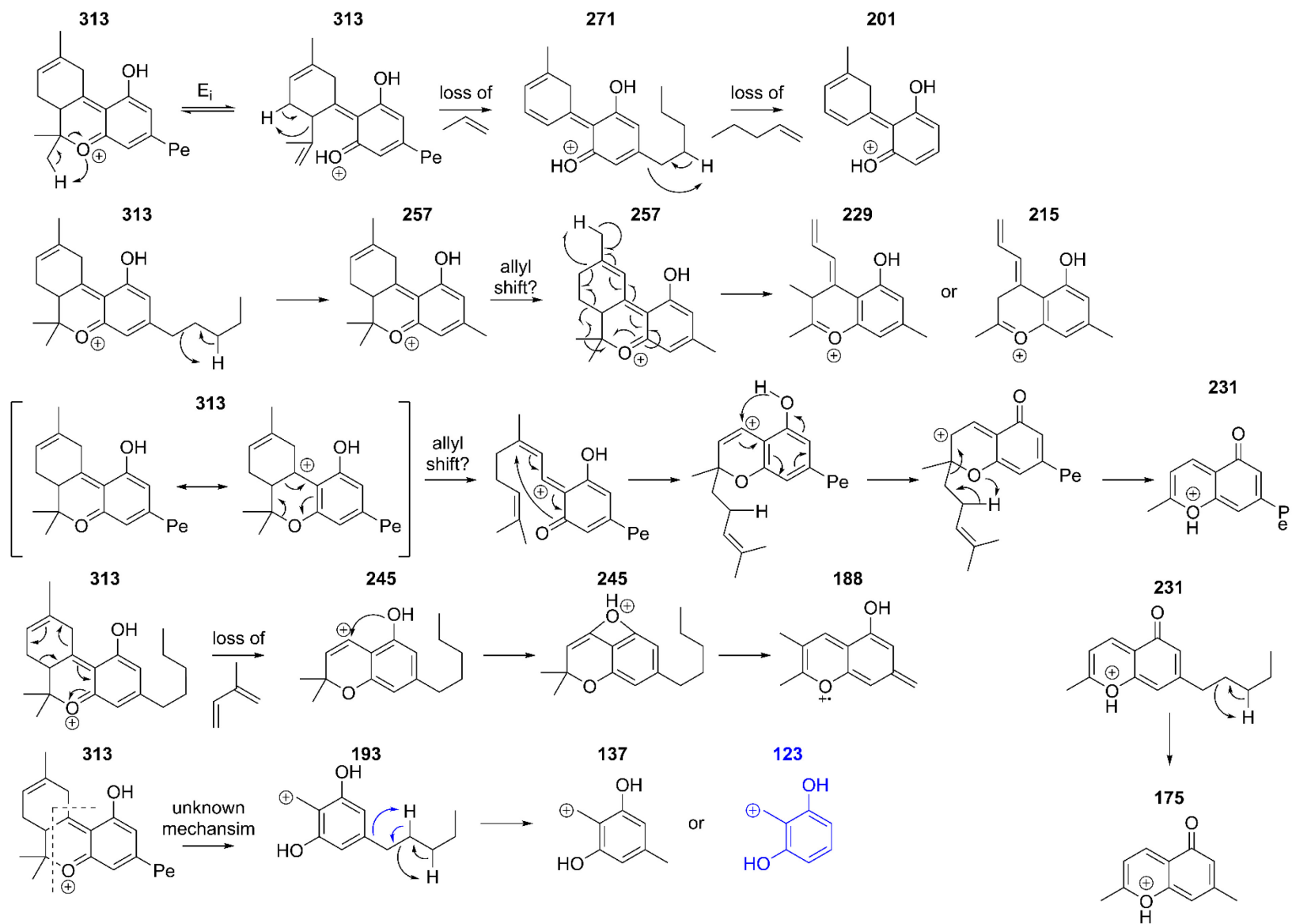


Figure S5-8. Proposed mechanisms of formation for fragments originating from  $[\Delta^8\text{-THC} + \text{Ag}]^+$ .

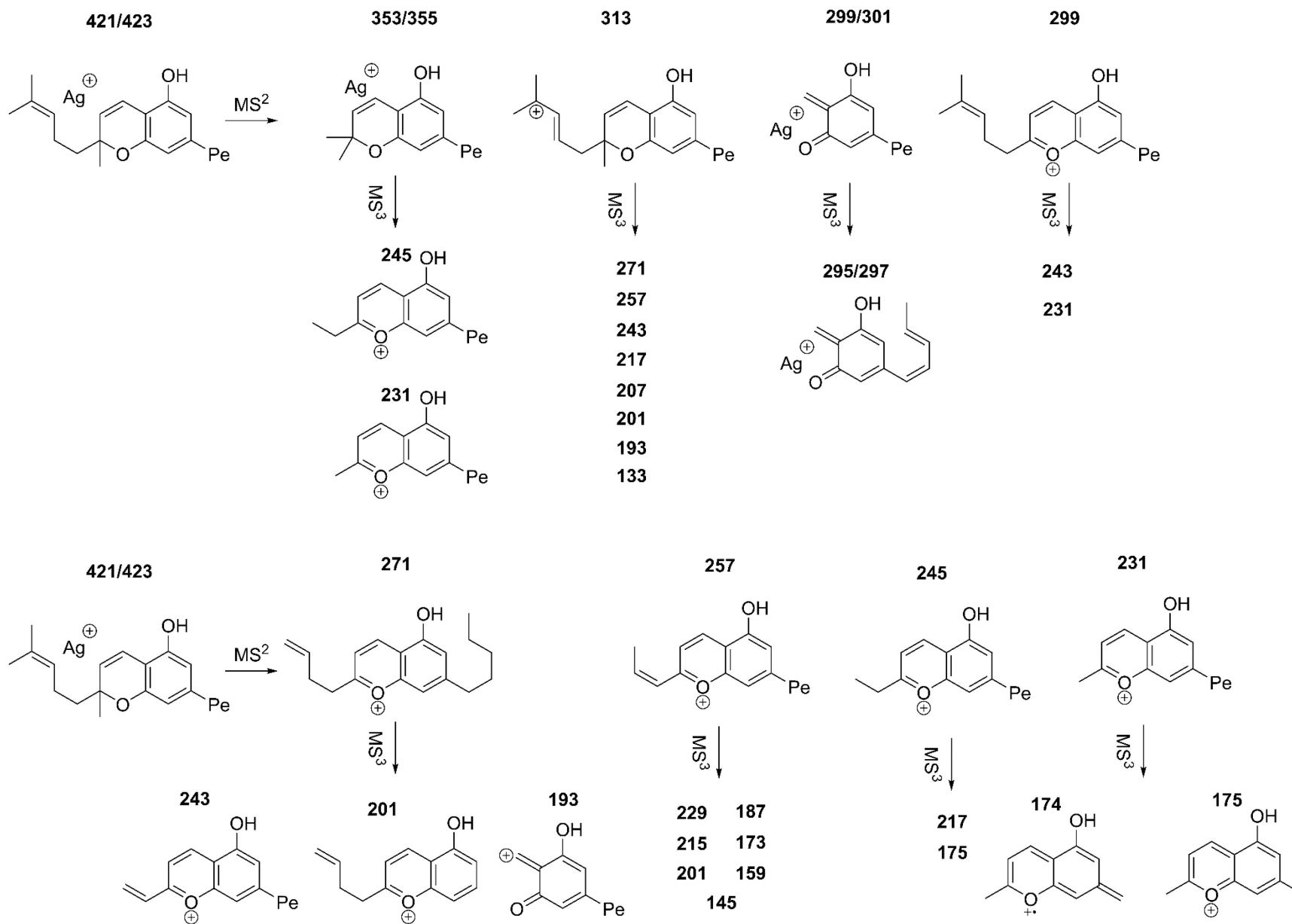


Figure S5-9. Proposed fragmentation pathway for [CBC + Ag]<sup>+</sup>.

Plausible mechanisms of formation for the most prominent fragments of [CBC + Ag]<sup>+</sup>

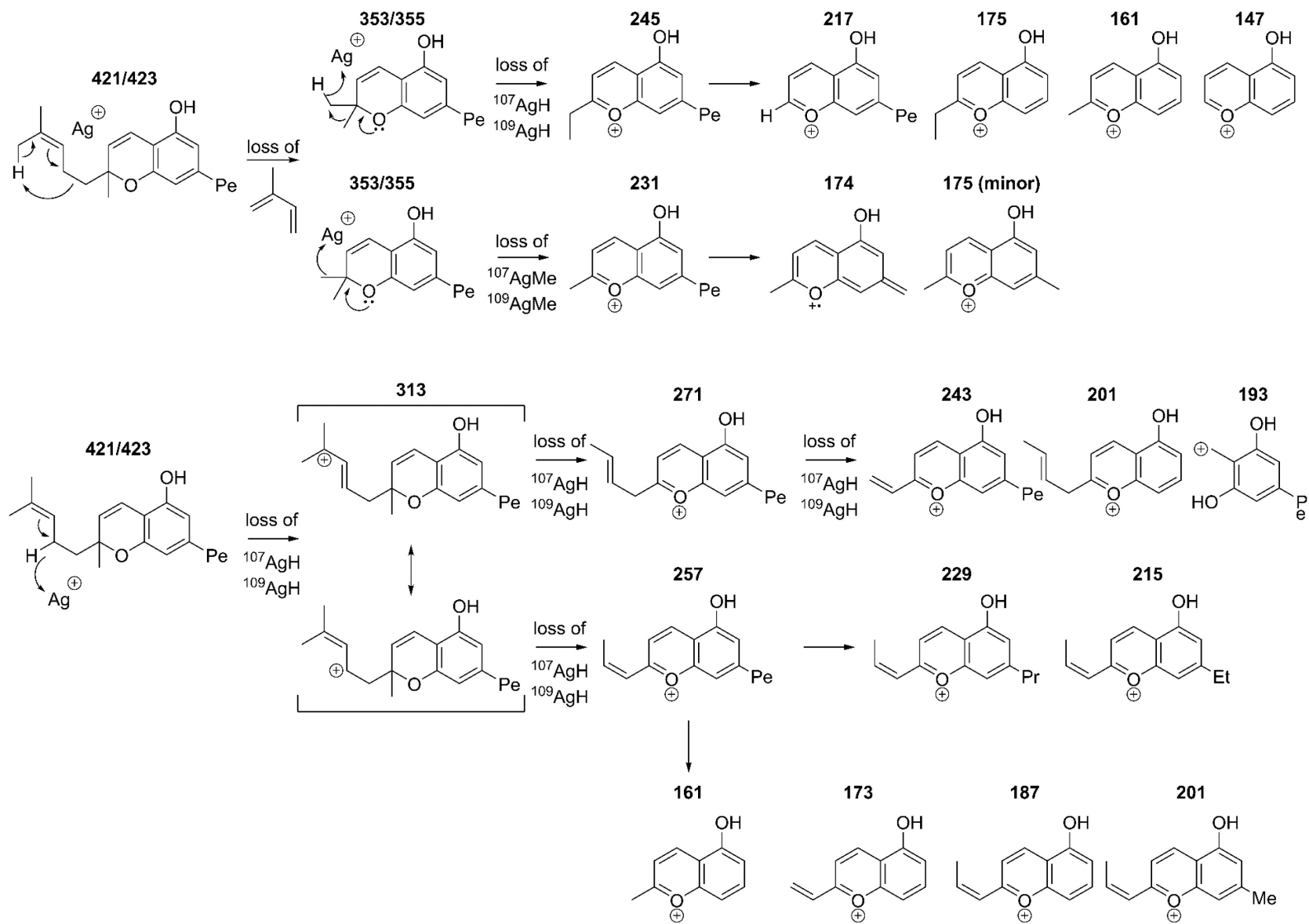
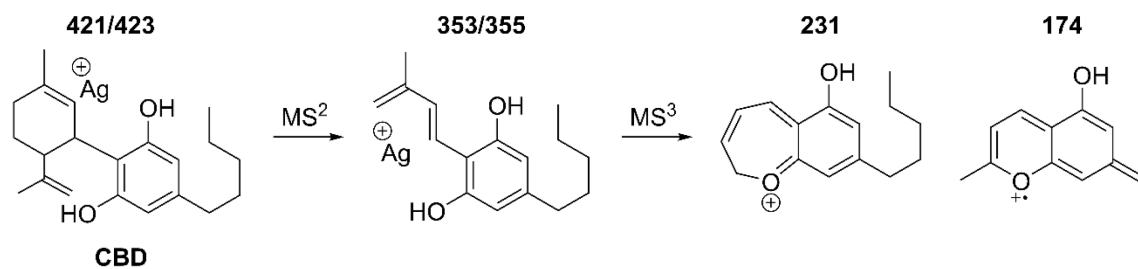
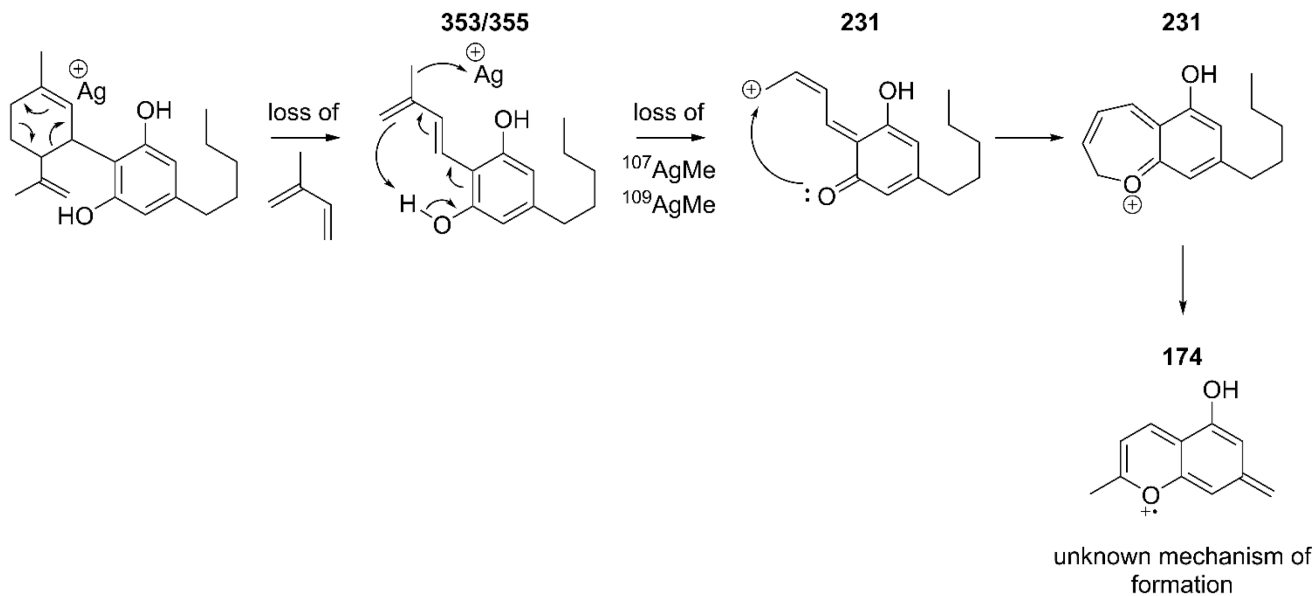


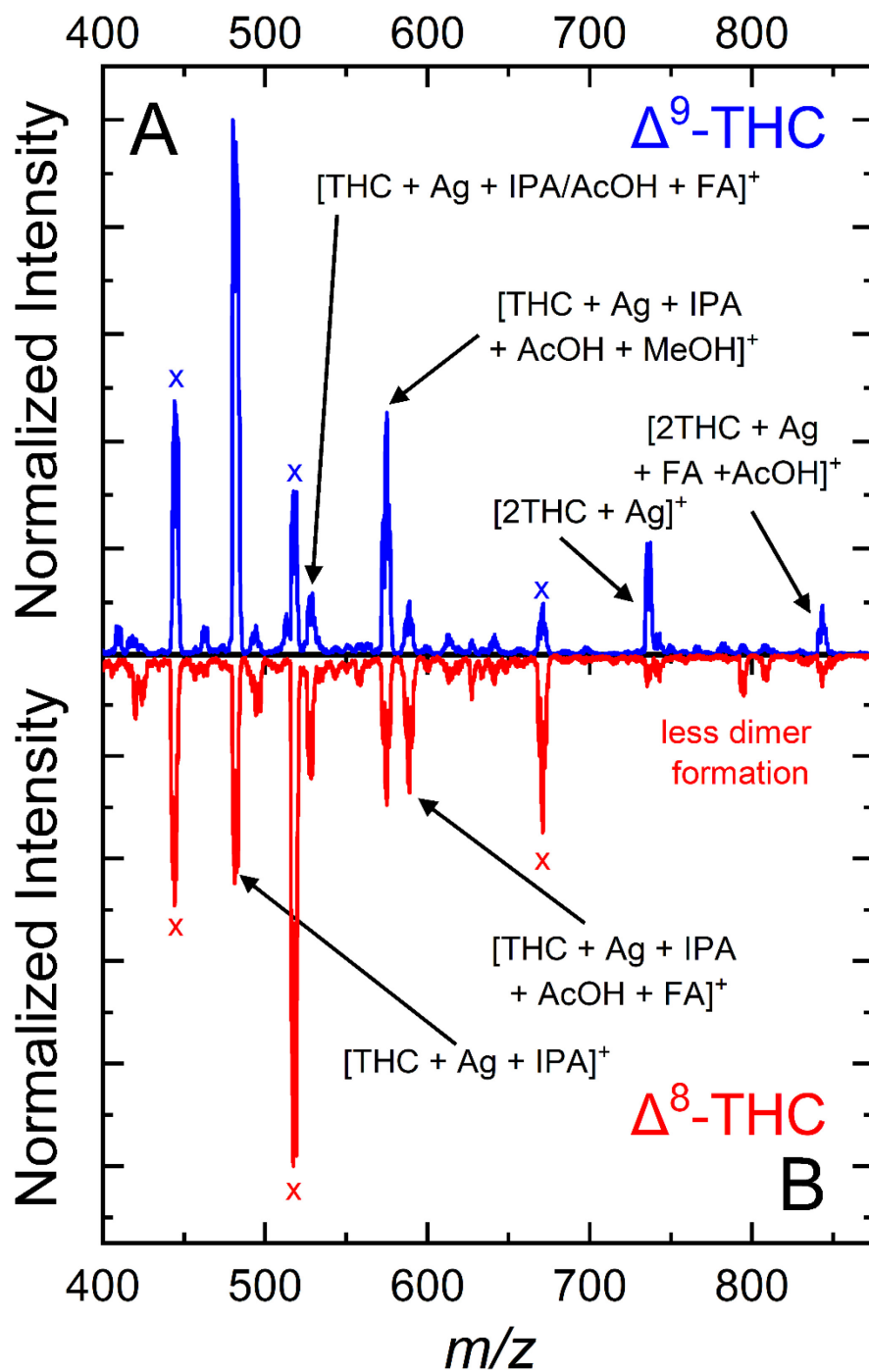
Figure S5-10. Proposed mechanisms of formation for fragments originating from [CBC + Ag]<sup>+</sup>.



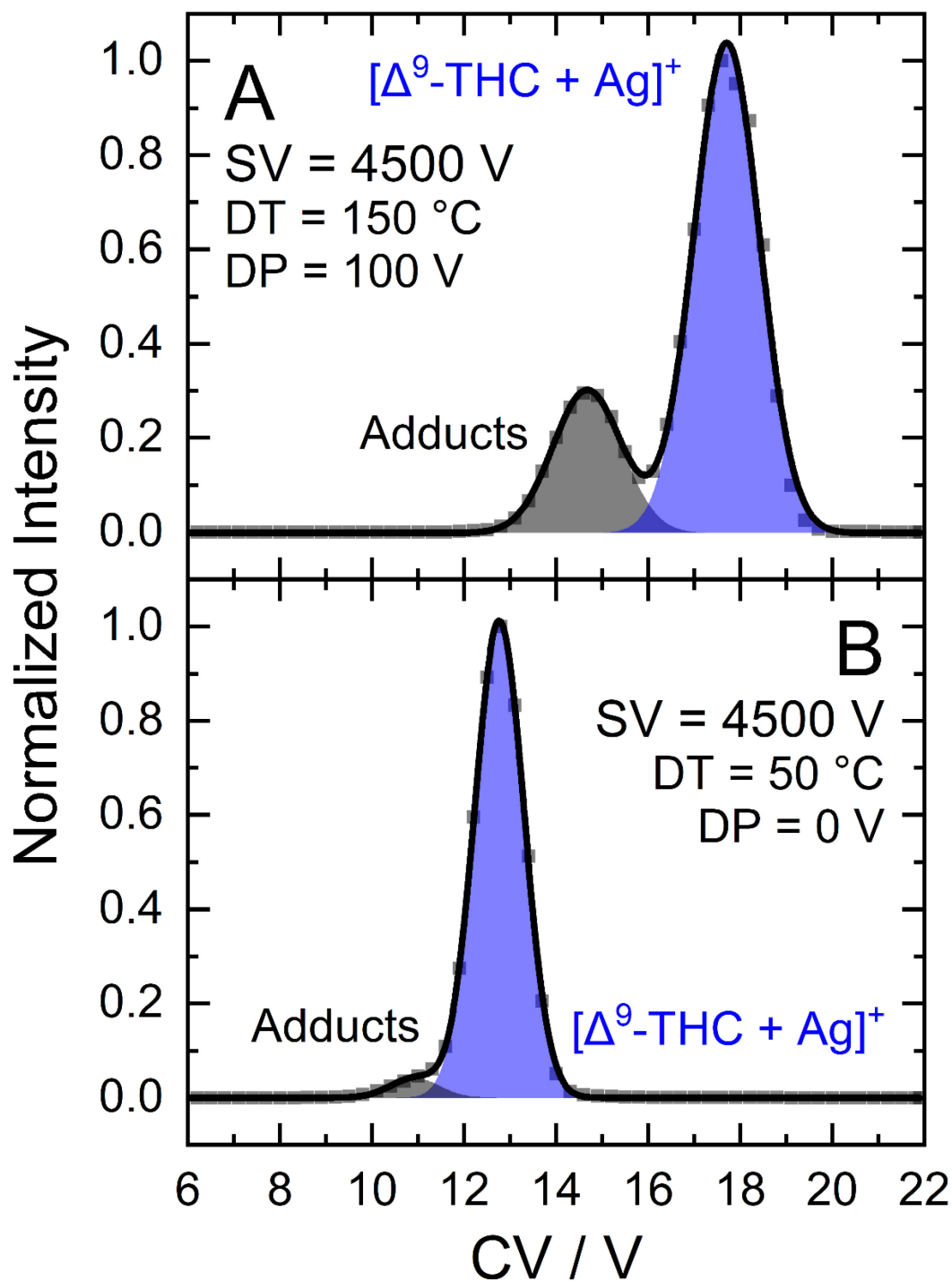
Plausible mechanisms of formation:



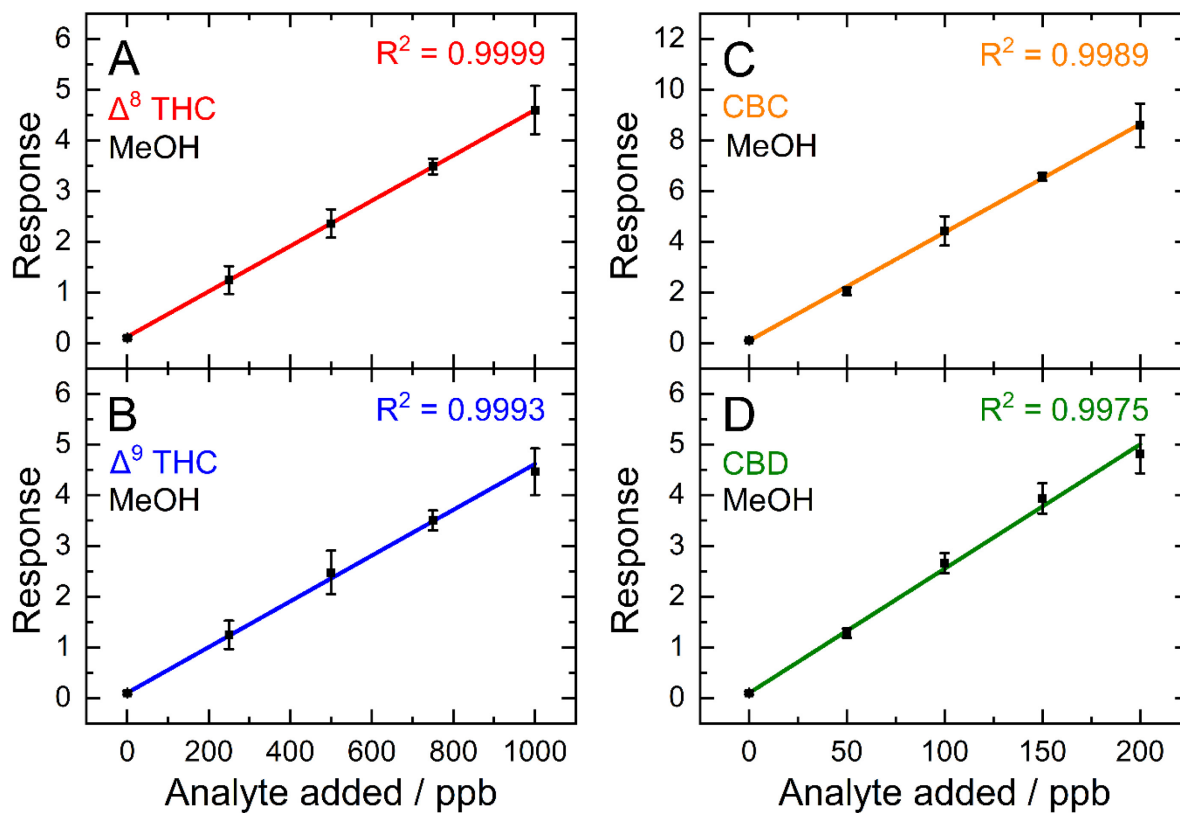
**Figure S5-11.** Proposed fragmentation pathway and mechanisms of formation for fragments originating from [CBD + Ag]<sup>+</sup>.



**Figure S5-12.** Q1 full scan mass spectrum of (A)  $[\Delta^9\text{-THC} + \text{Ag}]^+$  and (B)  $[\Delta^8\text{-THC} + \text{Ag}]^+$  produced via ESI from  $\text{H}_2\text{O}/\text{MeOH}$  (50:50 w/ 0.1%  $\text{HCOOH}$ ) containing 1 ppm of  $\Delta^8\text{-THC}/\Delta^9\text{-THC}$  and 5 ppm of  $\text{AgOAc}$ . 1.5 mol% of IPA was seeded into the carrier gas ( $\text{N}_2$ ), causing formation of ion-solvent clusters. Note that  $\text{AcOH}$  = acetic acid, IPA = isopropyl alcohol, FA = formic acid, and x denotes a species that does not contain  $\Delta^8\text{-THC}/\Delta^9\text{-THC}$ .

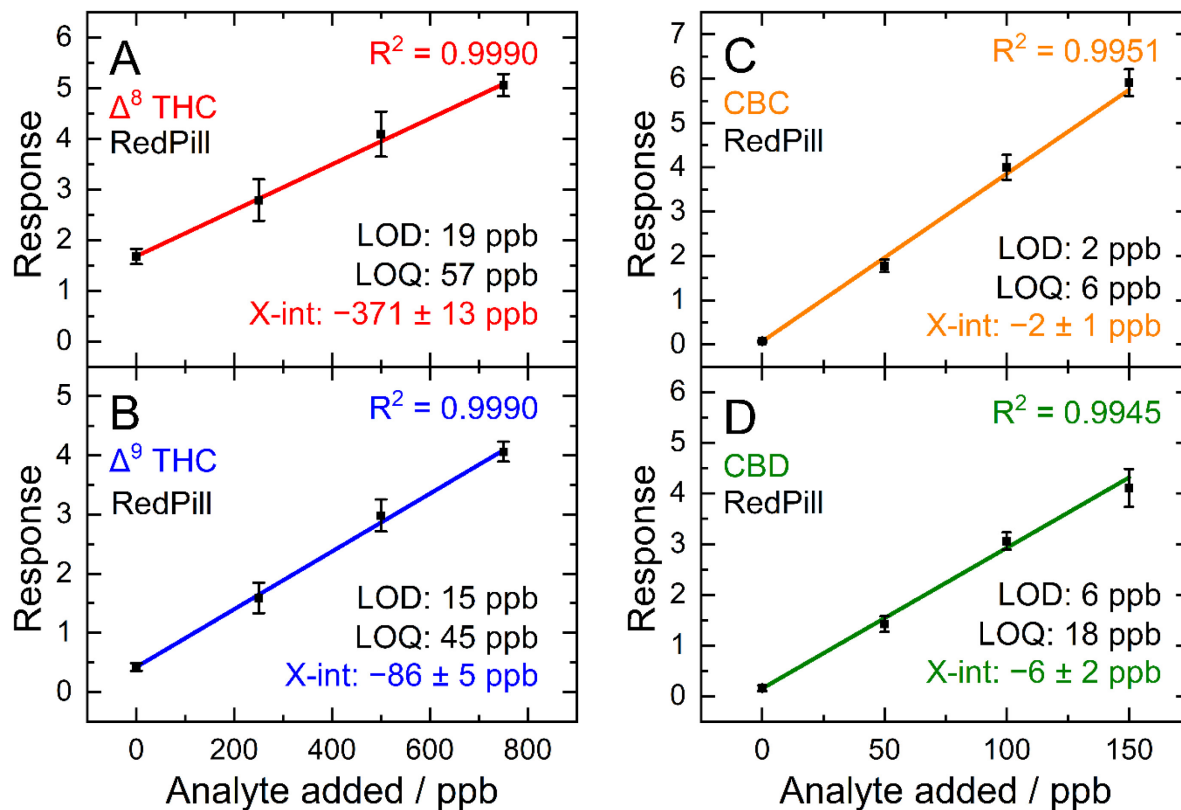


**Figure S5-13.** Ionogram of  $[\Delta^9\text{-THC} + \text{Ag}]^+$  obtained at (A) SV = 4500 V, DT = 150 °C, and DP = 100 V, or (B) SV = 4500 V, DT = 50 °C, and DP = 0 V. Reducing DT and DP mitigates the dissociation of the  $[\Delta^9\text{-THC} + \text{Ag} + \text{AcOH}]^+$  and  $[\Delta^9\text{-THC} + \text{Ag} + \text{AcOH} + \text{MeOH}]^+$  adducts that give rise to the additional peak in panel A.

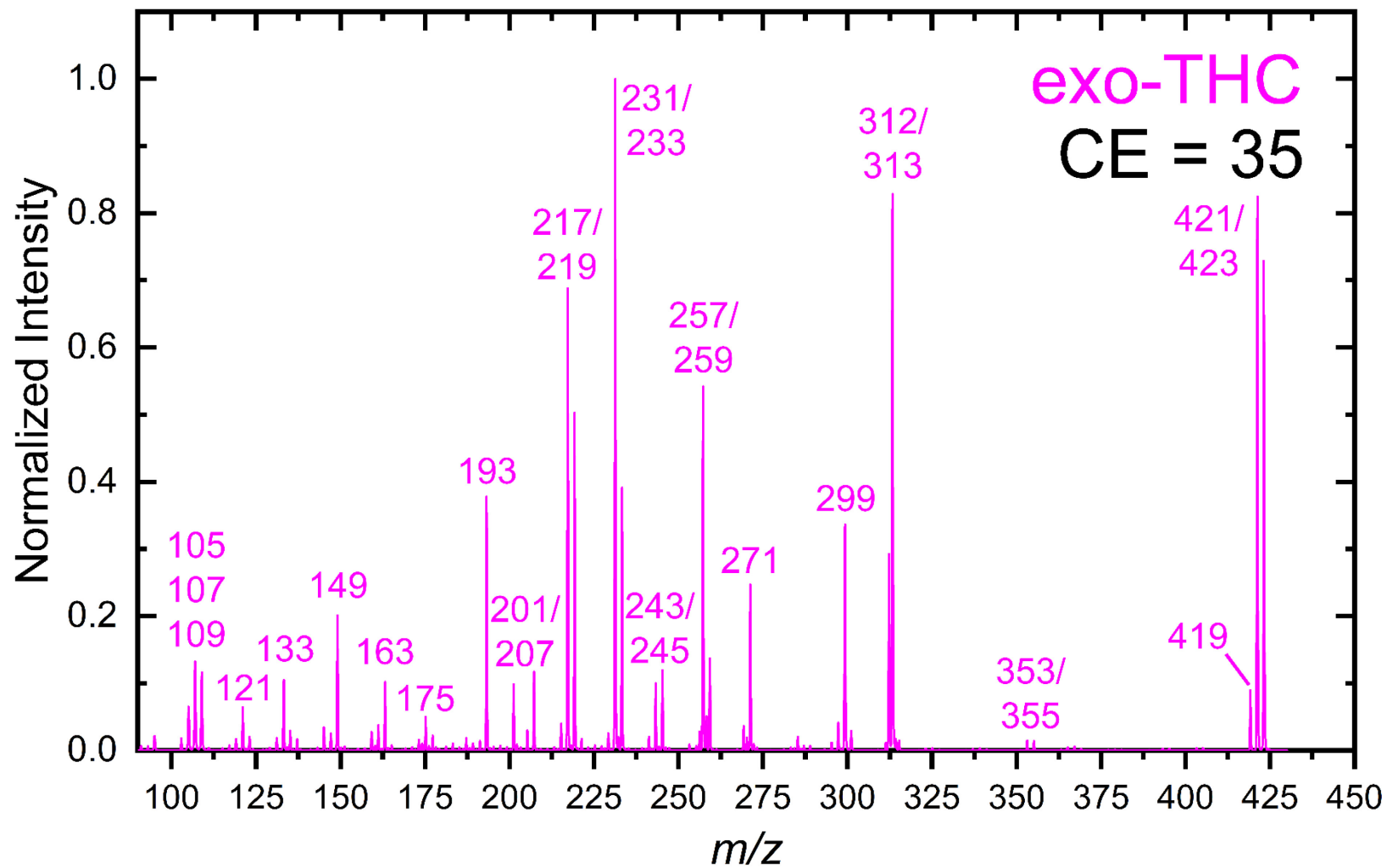


**Figure S5-14.** Calibration curves from concurrent standard additions of (A)  $\Delta^8$ -THC, (B)  $\Delta^9$ -THC, (C) CBC, and (D) CBD to a solution of MeOH containing 5 ppm of AgOAc. Error bars correspond to the standard deviation of the analyte response relative to the internal standard (CBDA; 200 ppb) across three replicate measurements.

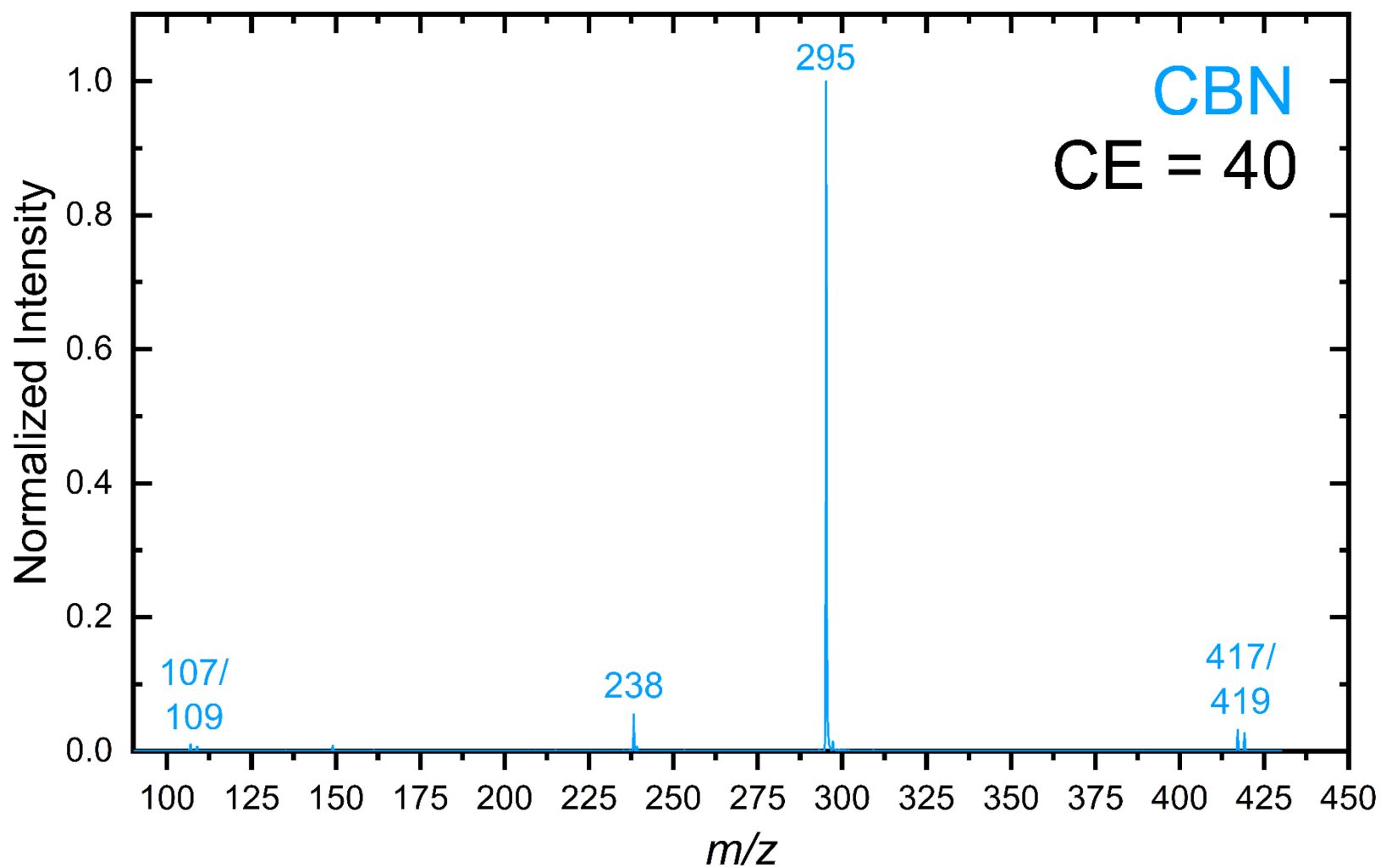




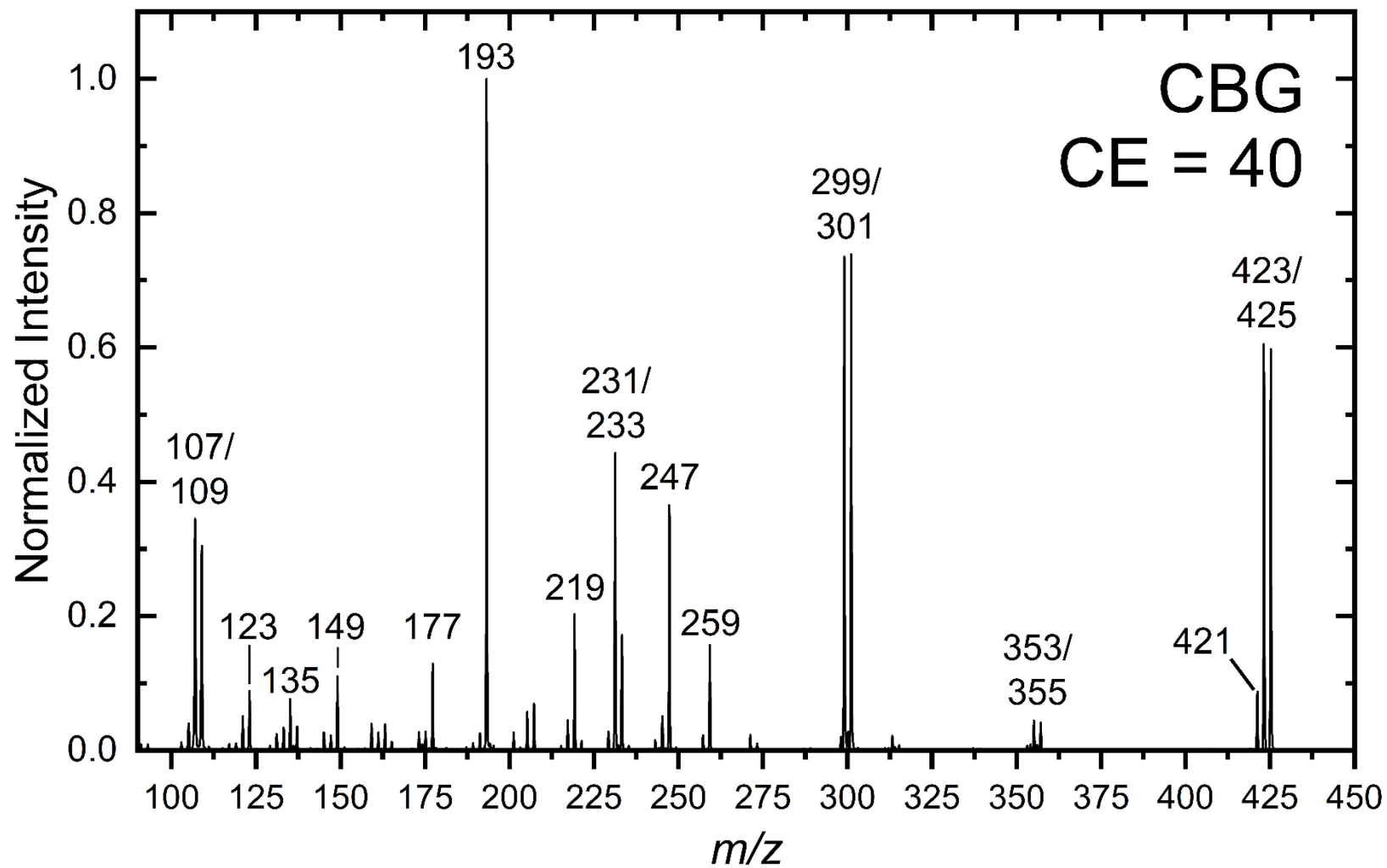
**Figure S5-15.** Calibration curves from concurrent standard additions of (A)  $\Delta^8$ -THC, (B)  $\Delta^9$ -THC, (C) CBC, and (D) CBD to the RedPill extract (50000-fold dilution in MeOH and doped with 5 ppm of AgOAc). Error bars correspond to the standard deviation of the analyte response relative to the internal standard (CBDA; 200 ppb) across three replicate measurements. Limits of detection (LOD) and limits of quantitation (LOQ) are using Equations 4 and 5 (see main text).



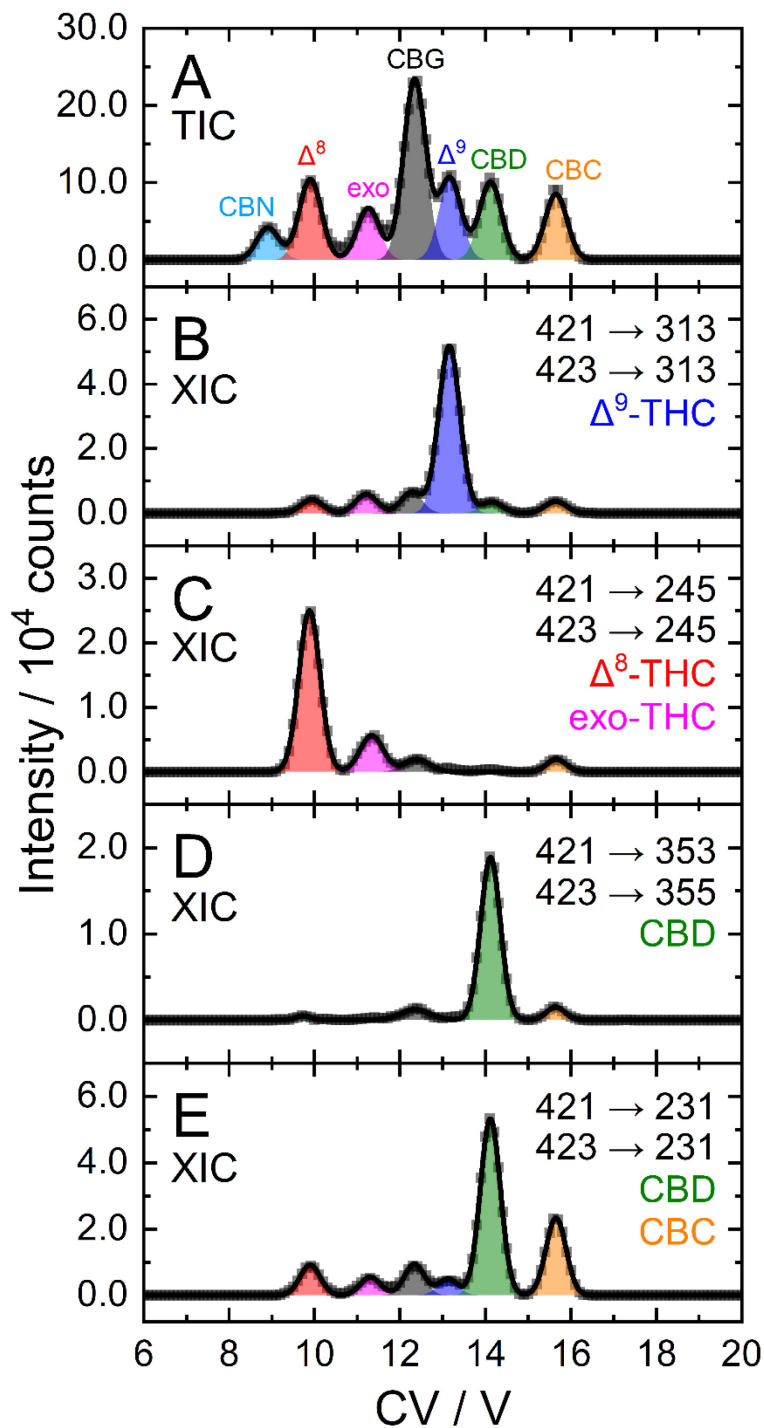
**Figure S5-16.** Product-ion spectrum of [exo-THC + Ag]<sup>+</sup> at CE = 35. Note that Q1 was modified to transmit both *m/z* 421 and 423 isotopologues of [exo-THC + Ag]<sup>+</sup>.



**Figure S5-17.** Product-ion spectrum of  $[\text{CBN} + \text{Ag}]^+$  at  $\text{CE} = 40$ . Note that Q1 was modified to transmit both  $m/z$  417 and 419 isotopologues of  $[\text{CBN} + \text{Ag}]^+$ .



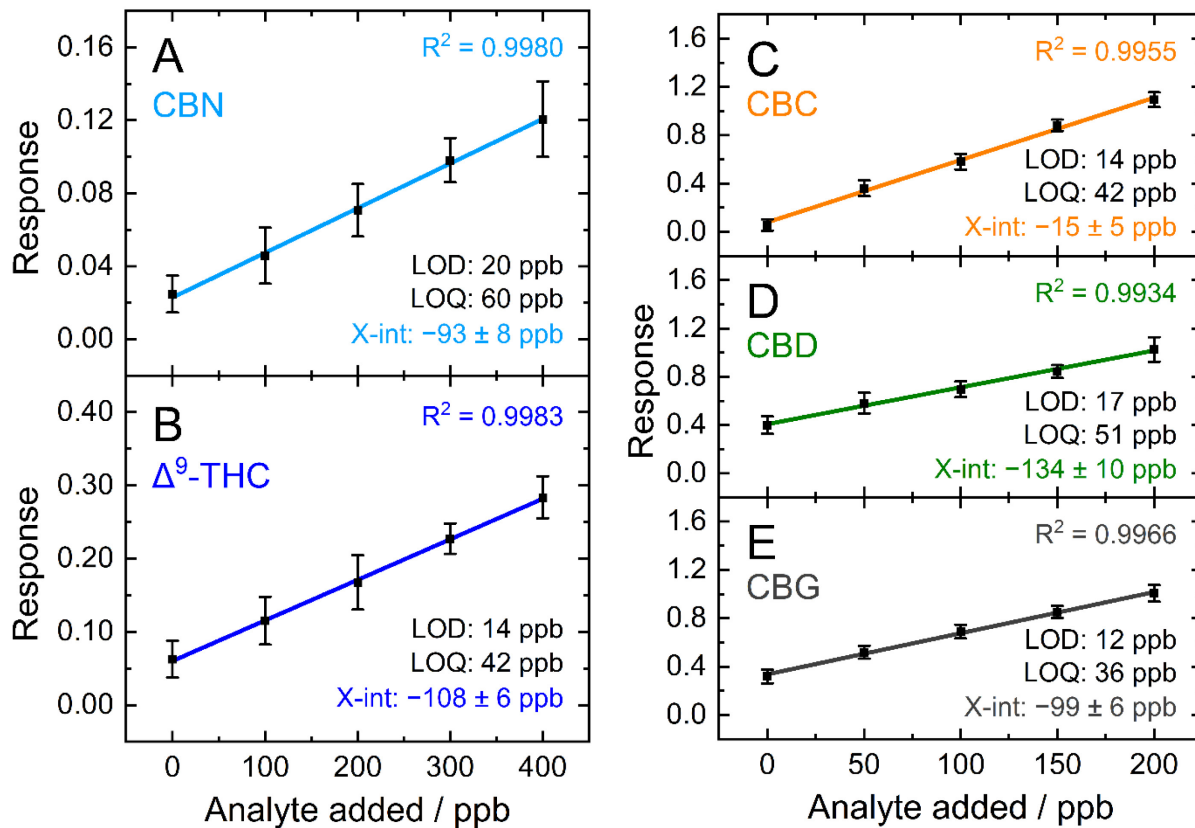
**Figure S5-18.** Product-ion spectrum of  $[\text{CBG} + \text{Ag}]^+$  at  $\text{CE} = 40$ . Note that Q1 was modified to transmit both  $m/z$  423 and 425 isotopologues of  $[\text{CBG} + \text{Ag}]^+$ .



**Figure S5-19.** (A) Total ion chromatogram (TIC) and (B – E) extracted ion chromatograms (XIC) from a specific MRM transition for the separation of argentinated  $\Delta^8$ -THC (red),  $\Delta^9$ -THC (blue), CBC (orange), CBD (green), exo-THC (magenta), CBN (cyan), and CBG (black) by DMS-MS. Ionograms were obtained in a pure N<sub>2</sub> environment at SV = 4500 V, DP = 0 V, and DT = 50 °C from a mixture containing  $\Delta^8$ -THC (500 ppb),  $\Delta^9$ -THC (500 ppb), CBD (100 ppb), CBC (100 ppb), exo-THC (500 ppb), CBN (500 ppb), CBG (100 ppb), and AgOAc (10 ppm) in MeOH.



**Figure S5-20.** Packaging of the Full Spectrum Hemp Multicannabinoid Oil extract produced by NuLeaf Naturals. At the top, the concentration of CBN,  $\Delta^9$ -THC, CBG, CBD, and CBC is stated to be  $12 \text{ mg mL}^{-1}$ , yet below, the amount of cannabinoid per activation (1 activation =  $0.5 \text{ mL}$ ) leads to concentrations of  $12.0 \text{ mg mL}^{-1}$  for CBN, CBG, CBD, and CBC, but  $13.6$  for  $\Delta^9$ -THC. This is also reflected in the  $\text{mg g}^{-1}$  equivalent, which we assume is determined from the density of the oil.



**Figure S5-21.** Calibration curves from concurrent standard additions of (A) CBN, (B)  $\Delta^9$ -THC, (C) CBC, and (D) CBD, and (E) CBG to the NuLeaf extract (133333.3-fold dilution in MeOH and doped with 10 ppm of AgOAc). Error bars correspond to the standard deviation of the analyte response relative to the internal standard (CBDA; 300 ppb) across three replicate measurements. Limits of detection (LOD) and limits of quantitation (LOQ) are using Equations 4 and 5 (see main text).

**Table S5-1.** MRM transitions of the deprotonated forms of  $\Delta^8$ -THC,  $\Delta^9$ -THC, CBD, and CBC [*i.e.*,  $M - H$ ]<sup>-</sup>, along with the cannabinoids that appear within the extracted ion chromatogram (XIC) upon separation by DMS-MS.

Q1	Q3	CE / V	Cannabinoids present in XIC
313	313	17	$\Delta^8$ -THC, $\Delta^9$ -THC, CBD, CBC
313	245	10	$\Delta^8$ -THC, $\Delta^9$ -THC, CBD
313	191	24	$\Delta^8$ -THC, $\Delta^9$ -THC, CBD, CBC
313	179	24	$\Delta^8$ -THC, $\Delta^9$ -THC, CBD, CBC

**Table S5-2.** MRM transitions of the argintinated cannabinoids  $[M + Ag]$ <sup>+</sup>. Cannabinoids in bold are the major peak in the extracted ion chromatogram (XIC), whereas cannabinoids in brackets are present in small quantities in the XIC (< 4 %).

Q1	Q3	CE / V	Cannabinoids present in XIC
421/423	313	17	<b><math>\Delta^9</math>-THC</b> ( $\Delta^8$ -THC, <i>exo</i> -THC, CBD, CBC)
421/423	245	10	<b><math>\Delta^8</math>-THC</b> ( <i>exo</i> -THC, CBG, $\Delta^9$ -THC, CBD, CBC)
421/423	353	24	<b>CBD</b> ( $\Delta^8$ -THC, <i>exo</i> -THC, CBG, $\Delta^9$ -THC, CBD, CBC)
421/423	355	24	<b>CBD</b> ( $\Delta^8$ -THC, <i>exo</i> -THC, CBG, $\Delta^9$ -THC, CBD, CBC)
421/423	231		<b>CBD, CBC</b> ( $\Delta^8$ -THC, <i>exo</i> -THC, CBG, $\Delta^9$ -THC)
421/423	219		<b>Exo-THC</b> ( $\Delta^8$ -THC, $\Delta^9$ -THC, CBC)
417/419	295		<b>CBN</b>
423/425	301		<b>CBG</b> ( $\Delta^8$ -THC, <i>exo</i> -THC, $\Delta^9$ -THC, CBD, CBC)
423/425	193		<b>CBG</b> ( $\Delta^8$ -THC, <i>exo</i> -THC, $\Delta^9$ -THC, CBD, CBC)



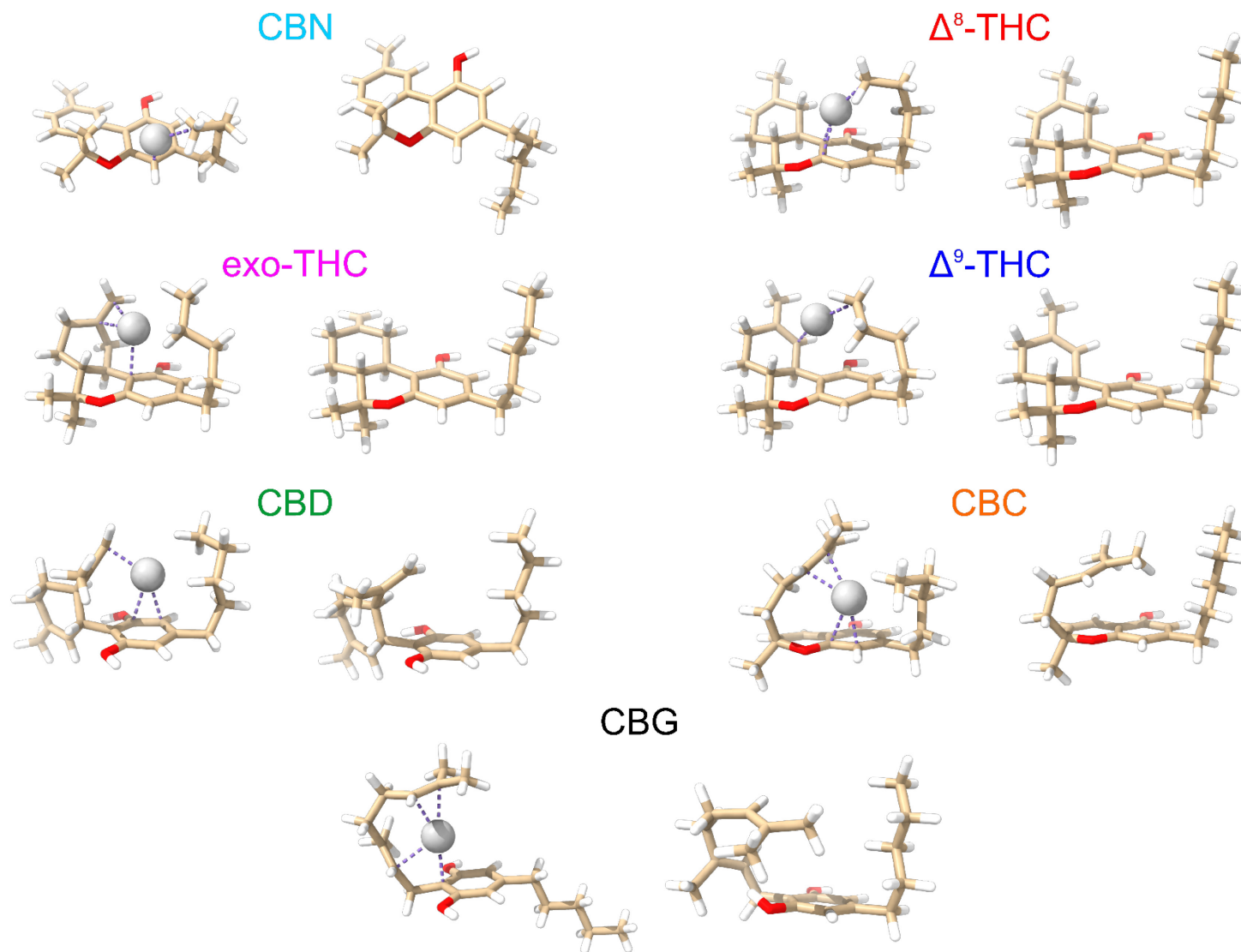
**Table S5-3.** Operating conditions of the ESI-DMS-QTRAP5500.

Parameter	Setting
<b>Source</b>	
Curtain Gas (CUR)	20.0
Collision gas (CAD)	High (12.0)
Ion spray voltage (IS)	5500
Source Temperature (TEM)	50
Nebulizing Gas Pressure (GS1)	30.0
Auxiliary Gas Pressure (GS2)	20.0
<b>Compound</b>	
Declustering Potential (DP)	100
Entrance Potential (EP)	10
Collision Energy (CE)	Varies
Collision Cell Exit Potential (CXP)	15
<b>DMS</b>	
DMS Temperature (DT)	50, 150, 225, 300
Modifier (MD)	Pure N <sub>2</sub> or N <sub>2</sub> with 1.5 mol% IPA
Separation Voltage (SV)	4400 V
Compensation Voltage (CV)	Varies
DMS Offset (DMO)	-3.0
DMS Resolution Enhancement (DR)	Varies (0 – 35 psi)
<b>Resolution – Quad 1</b>	
Ion Energy 1 (IE1)	1.1
Q1 Resolution	Unit
<b>Resolution – Quad 3</b>	
Ion Energy 3 (IE3)	1.8
Q3 Resolution	Unit
<b>Detector</b>	
CEM (CEM)	2100.0

## Supplementary Sections

### Section 5-1. Computational Methods

Quantum-chemical calculations were performed to assess the affinity of each cannabinoid for argentation. Briefly, candidate geometries for the neutral and argentated form of  $\Delta^8$ -THC,  $\Delta^9$ -THC, CBD, CBC, cannabiol (CBN), cannabigerol (CBG), and exo-THC were generated manually. Each candidate structure was used as an input for the Conformer-Rotamer Ensemble Sampling Tool (CREST),<sup>1,2</sup> which generated a series of low-energy structures using the GFN2-xTB semiempirical tight-binding model.<sup>3</sup> For each neutral and argentated cannabinoid, all structures generated by CREST that were within 25 kJ mol<sup>-1</sup> of the lowest energy conformer were extracted and additionally sorted by cosine similarity,<sup>4</sup> yielding 5 – 30 representative conformers. Each of the unique conformers was carried forward for optimization at the  $\omega$ B97X-D3/Def2-TZVPP level of theory, which employed the RIJcosX approximation and the Def2/J auxiliary basis set.<sup>5-9</sup> Normal mode analyses were conducted to ensure that each structure corresponded to a true minimum (*i.e.*, no imaginary frequencies) and to calculate thermochemical corrections. Electronic energies of the optimized structures were refined with single-point energy calculations conducted at the DLPNO-CCSD(T)/Def2-TZVPP level of theory, which used the Def2/C auxiliary basis set.<sup>10-13</sup> Thermochemical quantities were determined by combining DLPNO-CCSD(T) single point electronic energies with  $\omega$ B97X-D3 thermochemistry, which we report as DLPNO-CCSD(T)/Def2-TZVPP// $\omega$ B97X-D3/Def2-TZVPP. All *ab initio* and density functional theory (DFT) calculations were performed using the ORCA computational package (version 5.0.3).<sup>14-17</sup> Results of the quantum-chemical calculations are provided in the ioChem-BD database entry associated with this manuscript (<https://doi.org/10.19061/iochem-bd-6-225>).<sup>18</sup> For the convenience of the reader, the structures of the lowest energy configuration of each argentated and neutral cannabinoid are provided in Figure S5-22.



**Figure S5-22.** Lowest energy structure of the argentinated forms (i.e.,  $[M + Ag]^+$ ) and neutral forms of CBN,  $\Delta^8$ -THC, exo-THC,  $\Delta^9$ -THC, CBD, CBC, and CBG, as determined at the DLPNO-CCSD(T)/Def2-TZVPP// $\omega$ B97X-D3/Def2-TZVPP level of theory.

The affinity of each cannabinoid for argentation is determined by the Gibbs energy of association ( $\Delta G_{ass}$ ; Equation S5-1) , where  $G_{Ag^+}$ ,  $G_M$ , and  $G_{[M+Ag]^+}$  are the Gibbs energies of the silver cation, neutral cannabinoid, and argentinated cannabinoid, respectively. Calculation of  $\Delta G_{ass}$  uses weighted Gibbs energies, where the population  $\rho$  of the  $n^{th}$  isomer in an ensemble is determined by its Gibbs corrected electronic energy,  $G_n$ , (Equation S5-2; T = 298 K). The Gibbs energy of the ensemble,  $G$ , is given by the product of the  $n^{th}$  isomer's population and Gibbs energy, summed over all  $n$  isomers (Equation S5-3).

$$\Delta G_{ass} = (G_{Ag^+}) + (G_M) - (G_{[M+Ag]^+}) \quad Eq\ S5-1$$

$$\rho_n = \frac{\exp\left(\frac{G_n}{RT}\right)}{\sum_n \exp\left(\frac{G_n}{RT}\right)} \quad Eq\ S5-2$$

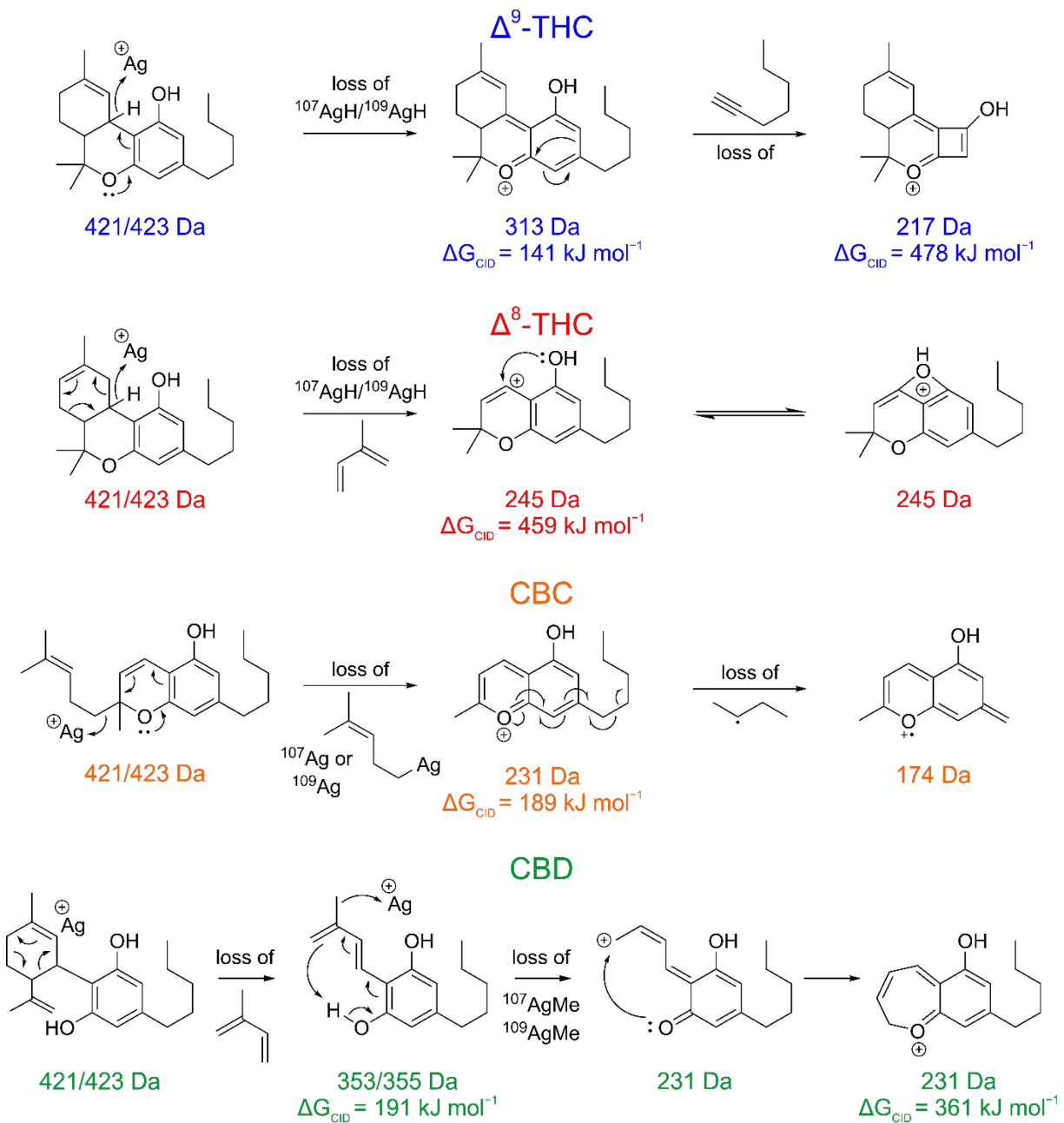
$$G = \sum_n \rho_n G_n \quad Eq\ S5-3$$

## Section S5-2. Computational investigation of the fragmentation behaviour of argentinated $\Delta^8$ -THC, $\Delta^9$ -THC, CBD, and CBC.

The distinct fragmentation patterns of the argentinated cannabinoid isomers is an intriguing observation, and one which is worthy of thorough investigation. Such an investigation would entail a detailed mapping of the reaction profile of each fragmentation path inclusive of transition states, as the kinetics of fragmentation during CID is especially important in rationalizing MS<sup>2</sup> behaviour. However, such a study is worthy of publication in its own right. Until such studies can be completed, we can gain insight into the MS<sup>2</sup> behaviour of each argentinated cannabinoid isobar by looking at the Gibbs energy associated with the proposed fragmentation pathways outlined in Figure S5-23 ( $\Delta G_{CID}$ ; Equation S5-4). Here,  $G_{Frag^+}$  is the Gibbs energy of the cationic fragment,  $G_{Frag}$  is the Gibbs energy of the neutral fragment, and  $G_{[M+Ag]^+}$  is the Gibbs energy of the intact, argentinated cannabinoid (*e.g.*,  $[\Delta^9\text{-THC} + \text{Ag}]^+$ ).

$$\Delta G_{CID} = (G_{Frag^+}) + (G_{Frag}) - (G_{[M+Ag]^+}) \quad \text{Eq S4}$$

Analysis of the  $\Delta G_{CID}$  values indicates that fragmentation pathways exhibit values that are in line with other computed values from previous computational studies.<sup>19</sup> Table S5-4 compares the  $\Delta G_{CID}$  for each possible product ion that originates from a particular argentinated precursor. For example, both  $[\Delta^9\text{-THC} + \text{Ag}]^+$  and  $[\Delta^8\text{-THC} + \text{Ag}]^+$  produce a fragment ion with a  $m/z$  of 313, except this fragment is only formed in small quantities from  $[\Delta^8\text{-THC} + \text{Ag}]^+$  but is the major fragment product from  $[\Delta^9\text{-THC} + \text{Ag}]^+$ . This is reflected in the lower  $\Delta G_{CID}$  value for the  $[\Delta^9\text{-THC} + \text{Ag}]^+$  fragmentation pathway relative to the fragmentation of  $[\Delta^8\text{-THC} + \text{Ag}]^+$ .



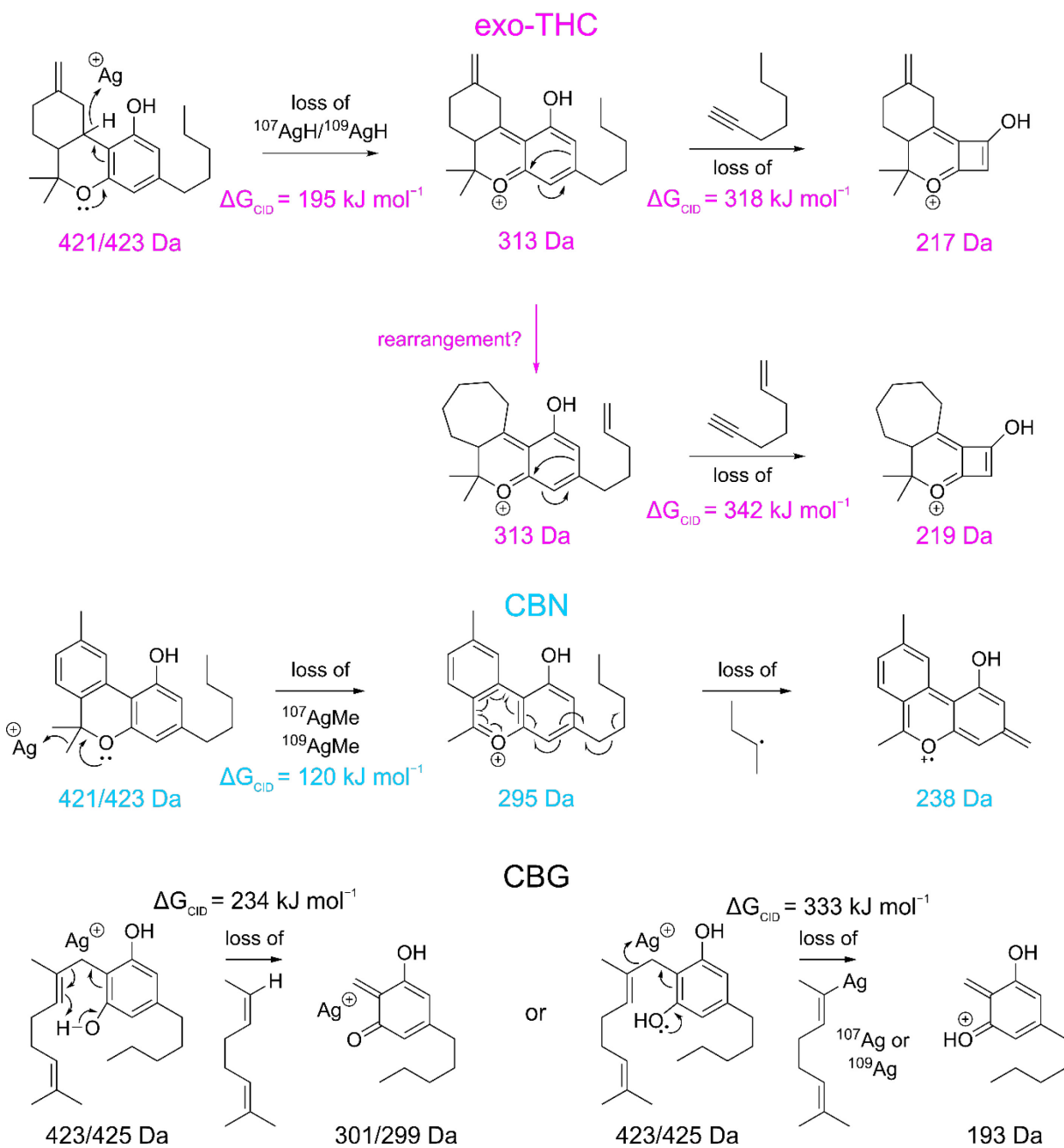
**Figure S23.** Proposed mechanisms of formation for the major product ions observed in the MS<sup>2</sup> spectra of [Δ<sup>9</sup>-THC + Ag]<sup>+</sup> (blue), [Δ<sup>8</sup>-THC + Ag]<sup>+</sup> (red), [CBC + Ag]<sup>+</sup> (orange), and [CBD + Ag]<sup>+</sup> (green).  $\Delta G_{\text{CID}}$  values are reported at the DLPNO-CCSD(T)/Def2-TZVPP// $\omega$ B97X-D3/Def2-TZVPP level of theory.

**Table S4.** Comparison of  $\Delta G_{CID}$  values (in  $\text{kJ mol}^{-1}$ ) for the fragments shown in Figure S5-23, as calculated at the DLPNO-CCSD(T)/Def2-TZVPP// $\omega$ B97X-D3/Def2-TZVPP level of theory.

Cannabinoid	$m/z$ 353/355	$m/z$ 313	$m/z$ 245	$m/z$ 231
$[\Delta^9\text{-THC} + \text{Ag}]^+$	—	141.3	—	—
$[\Delta^8\text{-THC} + \text{Ag}]^+$	—	165.2	459.5	—
$[\text{CBC} + \text{Ag}]^+$	—	—	—	188.9
$[\text{CBD} + \text{Ag}]^+$	191.0	—	—	361.5

### Section S5-3. Computational investigation of the fragmentation behaviour of argented exo-THC, CBN, and CBG.

Analogously to the results presented in Supplementary Section S2,  $\Delta G_{CID}$  values were computed for the major fragments originating from CID of the argented forms of exo-THC, CBN, and CBG. The proposed mechanisms of the formation of each fragment ion, alongside its respective  $\Delta G_{CID}$ , are provided below.





## Section 5-4. Frequently Asked Questions

1. **Are there any potential ramifications to instrument performance associated with infusing ~ 5ppm for AgOAc for prolonged periods of time?**

Some caution should always be taken when working with salt adducts, especially silver salts. Our experience has found that AgOAc is preferable to AgNO<sub>3</sub> in terms of preventing Ag buildup within the source and DMS cell. In the context of this work, the concentration of AgOAc is kept reasonably low (5 ppm or less) and does not cause and prolonged contamination within our system. A simple wipe of the curtain plate with a KimWipe lightly dipped in 1:1 MeOH/H<sub>2</sub>O removes the small amounts of silver that get deposited, and a flow of 1:1 MeOH/H<sub>2</sub>O for ~ 10 minutes removes all traces of silver and the argentinated cannabinoids from the DMS-MS system.

## References

- (1) Grimme, S. Exploration of Chemical Compound, Conformer, and Reaction Space with Meta-Dynamics Simulations Based on Tight-Binding Quantum Chemical Calculations. *J Chem Theory Comput* **2019**, *15* (5), 2847–2862. <https://doi.org/10.1021/acs.jctc.9b00143>.
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